# PfuTurbo DNA Polymerase AD

**Catalog** #600255, 600257, and 600259 600255-12, Revision A.01

### **MATERIALS PROVIDED**

	Quantity		
Materials provided	Catalog #600255	Catalog #600257	Catalog #600259
PfuTurbo DNA Polymerase AD	100 U	500 U	1000 U
10× Cloned Pfu Reaction Buffer AD	1 ml	2 × 1 ml	4 × 1 ml

**Storage:** Store at -20°C upon receipt.

#### Introduction

PfuTurbo DNA polymerase AD\* features a reformulated buffer system for increased economy with the same high performance as the original Stratagene PfuTurbo DNA polymerase for robust, high-fidelity PCR. As with the original PfuTurbo DNA polymerase, PfuTurbo DNA polymerase and the exclusive thermostable ArchaeMaxx polymerase-enhancing factor¹ that enhances PCR product yields and increases target length capability without altering DNA replication fidelity. PfuTurbo DNA polymerase AD can be used to amplify complex genomic DNA targets up to 19 kb and vector targets up to 15 kb in length. PfuTurbo DNA polymerase AD exhibits the same low error rate as Pfu DNA polymerase of 1–3 errors per 106 bases, while Taq DNA polymerase exhibits an error rate of 8–9 errors per 106 bases in the same assay.²

### OPTIMIZATION PARAMETERS (50 µL REACTION VOLUME)

Parameter	Genomic DNA Targets ≤6 kb or Vector DNA Targets ≤10 kb	Genomic DNA Targets >6 kb or Vector DNA Targets >10 kb	cDNA Targets
Extension time	1 minute per kb	2 minutes per kb	2 minutes per kb
PfuTurbo DNA polymerase AD	2.5 U	5.0 U	5.0 U
Input template	50–100 ng genomic DNA; 100 pg–30 ng vector DNA	200–250 ng genomic DNA; 100 pg–30 ng vector DNA	1–2 μl cDNA (from cDNA synthesis reaction)
Primers (each)	100–200 ng (0.2–0.5 μM)	200 ng (0.5 μM)	100-200 ng (0.2-0.5 μM)
dNTP concentration	100–250 μM each dNTP	500 μM each dNTP	100–250 μM each dNTP
Final reaction buffer conc.	1.0×	1.5× for genomic DNA or 1.0× for vector DNA	1.0× (supplemented with 1 mM MgSO <sub>4</sub> )
Denaturing temperature	95°C	92°C	95°C
Extension temperature	72°C	68°C	68°C

### **PCR PROTOCOL**

The reaction conditions given here are for amplification of a typical single-copy chromosomal target of  $\leq$ 6 kb. See the *Optimization Parameters* section for guidelines on amplifying longer targets, vector DNA targets, or cDNA targets. The reaction conditions are for one reaction and must be adjusted for multiple samples. The final volume of each reaction is 50  $\mu$ l.

1. Add the components in order into sterile thin-walled PCR tubes while mixing gently.

# Reaction Mixture for a Typical Single-Copy Chromosomal Locus PCR Amplification (≤6 kb)

Component	Amount per reaction
Distilled water (dH <sub>2</sub> O)	40.6 µl
10× Cloned Pfu Reaction Buffer AD a	5.0 μl
dNTP mix (25 mM each dNTP)	0.4 μl
DNA template (100 ng/µl)	1.0 μl
Primer #1 (100 ng/μl)	1.0 μl <sup>b</sup>
Primer #2 (100 ng/μl)	1.0 μl <sup>b</sup>
PfuTurbo DNA Polymerase AD (2.5 U/μl)	1.0 μl <sup>c</sup>
Total reaction volume	50 μΙ

- a The  $10\times$  buffer provides a final  $1\times$  Mg<sup>2+</sup> concentration of 2 mM. When amplifying cDNA, add Mg<sup>2+</sup> to a final  $1\times$  concentration of 3 mM. (For example, Mg<sup>2+</sup> concentration may be adjusted to 3 mM in the final 50- $\mu$ l reaction volume by adding 2  $\mu$ l of a PCR-grade 25 mM MgSO<sub>4</sub> solution and reducing the amount of dH<sub>2</sub>O to 38.6  $\mu$ l.)
- b The recommended primer concentration of  $0.2-0.5~\mu M$  corresponds to 100-200~ng for a typical 18- to 25-mer oligonucleotide primer in a  $50-\mu I$  reaction volume.
- The amount of *PfuTurbo* DNA polymerase AD varies depending on the length of the PCR target. The standard amount for vector targets up to 10 kb and genomic targets up to 6 kb in length is 1 µl (2.5 U).
- 2. If the extension time is >30 minutes, overlay each reaction with ~50 μl of DNase-, RNase-, and protease-free mineral oil (Sigma, St. Louis, Missouri).

3. Perform PCR using optimized cycling conditions. Suggested cycling parameters for *PfuTurbo* DNA polymerase AD-based PCR are provided below. (Optimized cycling parameters are not necessarily transferable between thermal cyclers. Consult the instrument manufacturer's recommendations if further optimization of cycling parameters is required.) Analyze the PCR amplification products on a 0.7–1.0% (w/v) agarose gel.

# PCR Cycling Program for PfuTurbo DNA Polymerase AD Genomic DNA Targets ≤6 kb and Vector DNA Targets ≤10 kb

Segment	Number of cycles	Temperature	Duration–Single Block Thermocycler	Duration–Stratagene RoboCycler Thermocycler
1	1	95°C⁰	1 minute	1 minute
2	30	95°C	30 seconds	1 minute
		Primer $T_{\rm m}$ – 5°C	30 seconds	1 minute
		72°C	1 minute per kb	1 minute per kb
3	1	72°C	10 minutes	10 minutes

## Genomic DNA Targets >6 kb, Vector DNA Targets >10 kb, and cDNA Targets

Segment	Number of cycles	Temperature	Duration–Single Block Thermocycler	Duration–Stratagene RoboCycler Thermocycler
1	1	92°C <sup>b</sup>	2 minutes	1 minute
2	10	92°C <sup>b</sup>	10 seconds	1 minute
		Primer $T_{\rm m} - 5^{\circ}{\rm C}$	30 seconds	1 minute
		68°C	2 minutes per kb	2 minutes per kb
3	20	92°C	10 seconds	1 minute
		Primer $T_{\rm m} - 5^{\circ}{\rm C}$	30 seconds	1 minute
		68°C	2 minutes per kb plus 10 seconds per cycle	2 minutes per kb plus 10 seconds per cycle

<sup>&</sup>lt;sup>a</sup> Denaturing temperatures above 95°C are recommended only for GC-rich templates.

#### TROUBLESHOOTING AND APPLICATION NOTES

- Low yield: If PCR product yields are lower than expected, optimize the extension time, amount of DNA template (excess DNA can inhibit PCR), and amount of *PfuTurbo* DNA polymerase AD. Optimize the PCR program denaturation time (typically 30–60 seconds at 95°C is sufficient; prolonged denaturation steps may damage the template DNA) and the annealing temperature. Extraneous salts contributed by primer or template DNA solutions may inhibit the PCR reaction. Use high-quality, gel purified primers and highly-purified template DNA.
- **Multiple bands:** The annealing temperature may require optimization. Typically annealing temperatures will range between 55°C and 72°C. Try adding Stratagene Perfect Match PCR Enhancer to improve specificity. Redesign primers.
- PCR adjuncts and cosolvents: Including a cosolvent, such as 1–10% (v/v) DMSO or 5–20% glycerol, or an adjunct, such as 1–5% formamide or 0.01–1 U of Stratagene Perfect Match PCR Enhancer, in the PCR reaction may increase performance for some targets/cycling programs.
- PCR cloning: If generating PCR fragments for cloning applications, use the StrataClone Blunt PCR Cloning Kit (Catalog #240207) or another blunt PCR cloning strategy. *PfuTurbo* DNA polymerase AD does **not** exhibit terminal deoxynucleotidyltransferase (TdT) activity, which is characterized by the addition of nontemplate-directed nucleotide(s) at the 3′ end of PCR-generated fragments.

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### REFERENCES

- 1. Hogrefe, H. H., Hansen, C. J., Scott, B. R. and Nielson, K. B. (2002) Proc Natl Acad Sci U S A 99(2):596-601.
- 2. Cline, J., Braman, J. C. and Hogrefe, H. H. (1996) Nucleic Acids Res 24(18):3546-51.

#### ENDNOTES

\* U.S. Patent Nos. 7,045,328, 6,734,293, 6,489,150, 6,444,428, 6,183,997, 5,948,663, 5,866,395, 5,545,552 and patents pending.

### **MSDS INFORMATION**

The Material Safety Data Sheet (MSDS) information for Stratagene products is provided on the web at <a href="http://www.stratagene.com/MSDS/">http://www.stratagene.com/MSDS/</a>. Simply enter the catalog number to retrieve any associated MSDS's in a print-ready format. MSDS documents are not included with product shipments.

 $<sup>^{\</sup>rm b}$  For cDNA targets, use a denaturation temperature of 95°C.